

Speeded up 60X. PtK2 cells. Note the beautiful lamella, by which the cell undergoes its own actin-based motility.

Gram positive bacillus

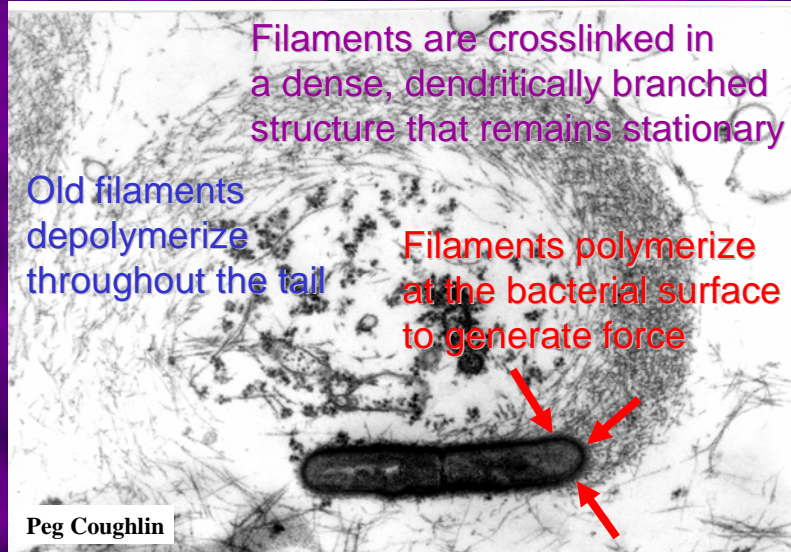
Propelled through the cytoplasm via actin polymerization

10 different labs working this out I do next step

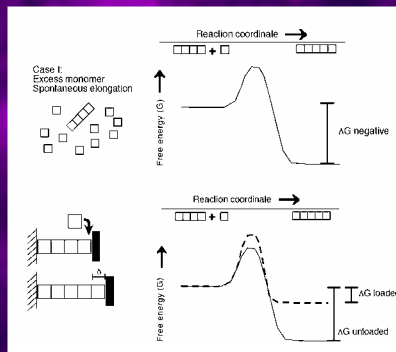
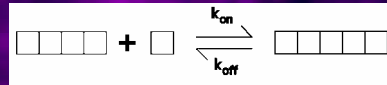
Other organisms which use this form of motility, beautiful example of coevolution. *Shigella*, *Rickettsia*, *Vaccinia*

Why evolve this motile mechanism? To run from the immune system.

## Bacterial surface proteins cause local nucleation of actin filaments

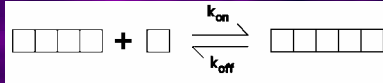


# Force generation by protein polymerization



Adapted from:  
 Hill & Kirschner, 1982  
 Int. Rev. Cytol. 78: 1-125

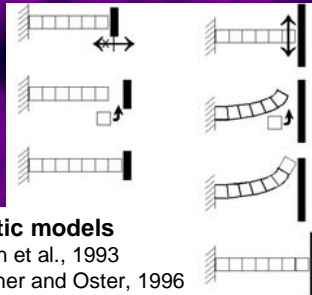
## Force generation by protein polymerization



$$F_{\max} = (kT/\delta) \ln(C/C_{\text{crit}})$$

$$C_{\text{crit}} \sim k_{\text{off}}/k_{\text{on}}$$

for actin:  $F_{\max} \sim 5\text{-}10 \text{ pN}$   
(comparable to myosin or kinesin)

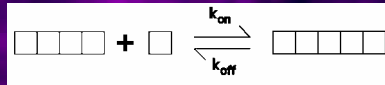


### Kinetic models

Peskin et al., 1993

Mogilner and Oster, 1996

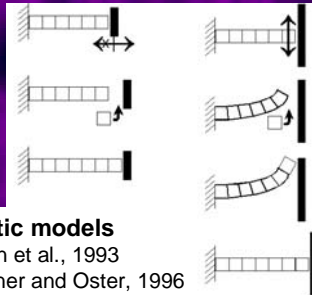
## Force generation by protein polymerization



$$F_{\max} = (kT/\delta) \ln(C/C_{\text{crit}})$$

$$C_{\text{crit}} \sim k_{\text{off}}/k_{\text{on}}$$

for actin:  $F_{\max} \sim 5\text{-}10 \text{ pN}$   
(comparable to myosin or kinesin)



### Kinetic models

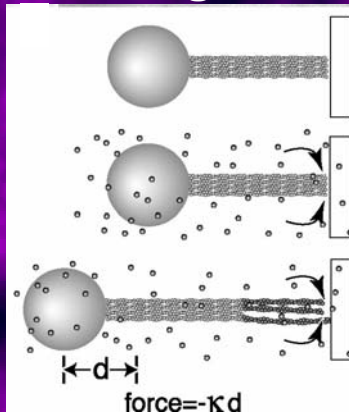
Peskin et al., 1993

Mogilner and Oster, 1996

How much force can be generated by growth of actin filaments against a rigid barrier?

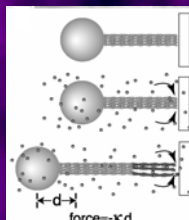
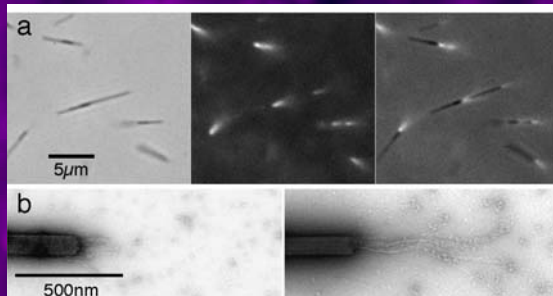
How fast can growth occur?

## Optical trap method for measuring force from growth of a small bundle



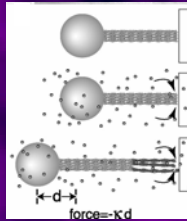
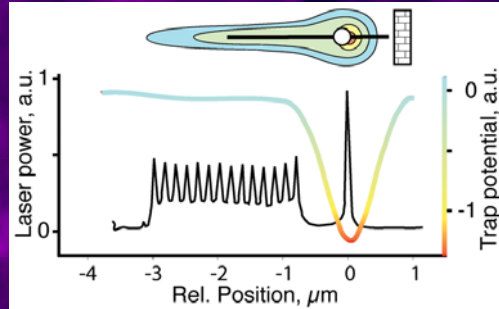
Collaboration with:  
Marileen Dogterom  
Matthew Footer  
Jacob Kerssemakers

## Optical trap method for measuring force from growth of a small bundle



Horseshoe crab sperm acrosomal bundles nucleate actin filament growth

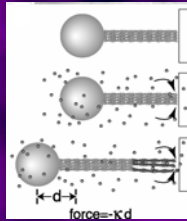
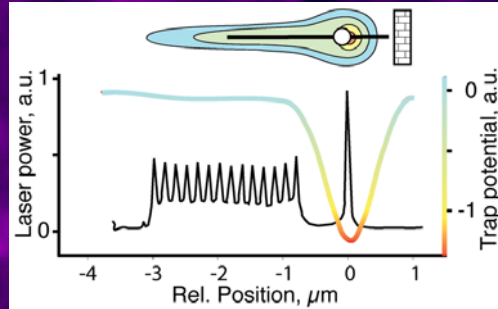
## Optical trap method for measuring force from growth of a small bundle



Keyhole optical trap holds bead and bundle next to wall

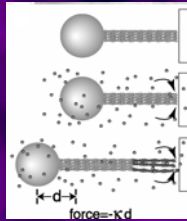
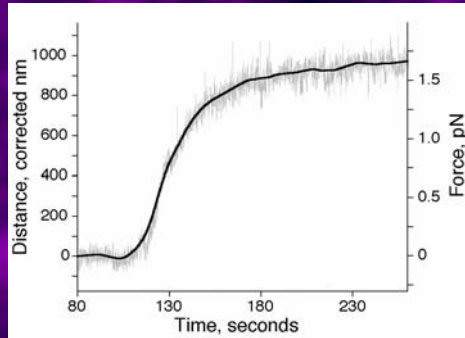


## Optical trap method for measuring force from growth of a small bundle



**Keyhole optical trap holds bead and bundle next to wall**

## Growth slows to stall at a few pN



Growth condition:  
 4  $\mu$ M actin  
 20  $\mu$ M profilin

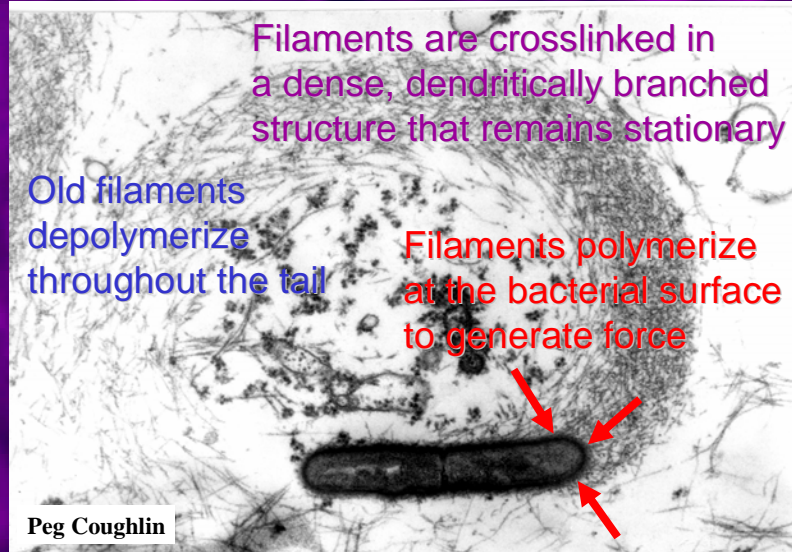
Stall occurs near  
 Hill & Kirschner limit

## Building up in scale

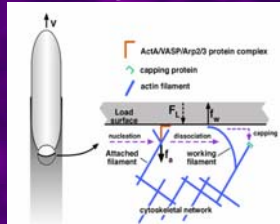
Individual filaments can generate a few picoNewtons of force; small bundles are not able to work together efficiently to push harder

Are branched networks better generators of polymerization-driven pushing force than bundles?

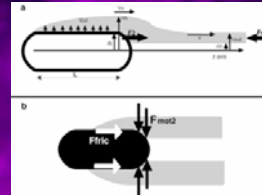
## Bacterial surface proteins cause local nucleation of actin filaments



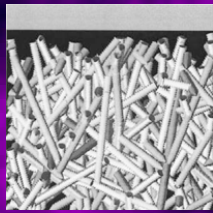
## Relationship between force and speed: Physical models at varying scales



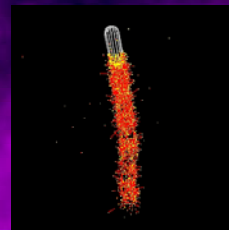
Microscopic:  
Mogilner and Oster, 2003



Mesoscopic:  
Gerbai, Chaikin, Rabin and Prost 2000

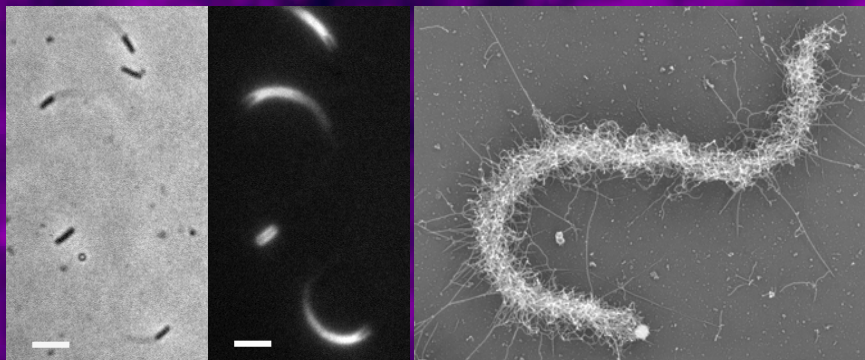


Bridging micro and meso:  
Carlsson, 2001



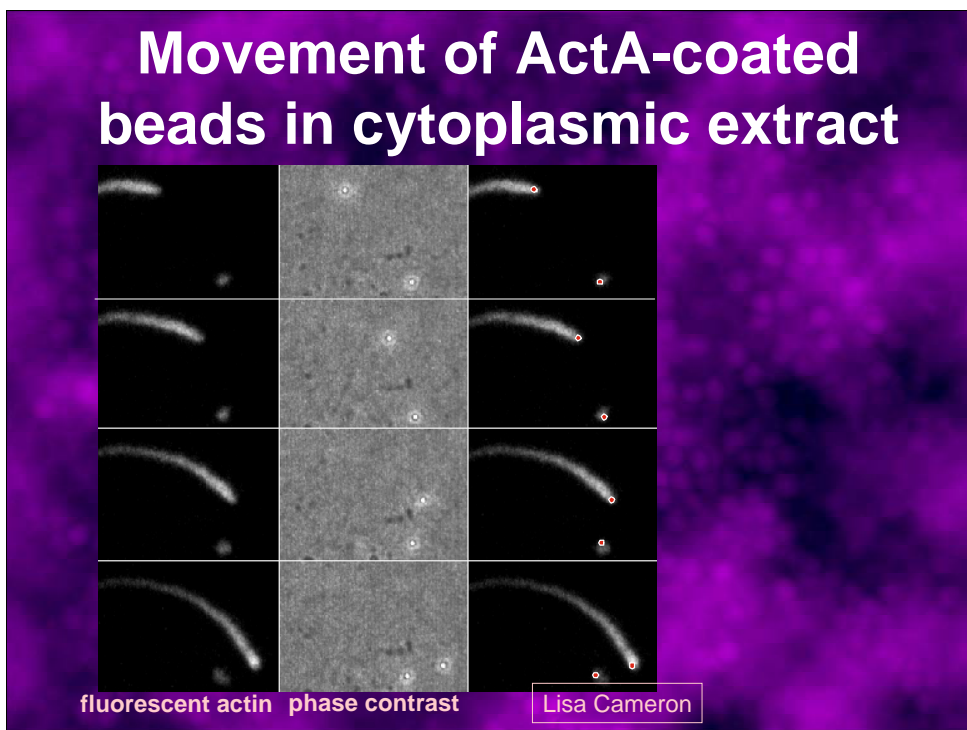
Alberts and Odell, 2004

## Biochemical and biophysical manipulations of actin comet tails



Movement in cytoplasmic extracts  
(Theriot et al., 1994)  
Reconstitution with purified proteins  
(Loisel et al., 1999)

Replacement of bacteria by  
ActA-coated polystyrene beads  
(Cameron et al., 1999)

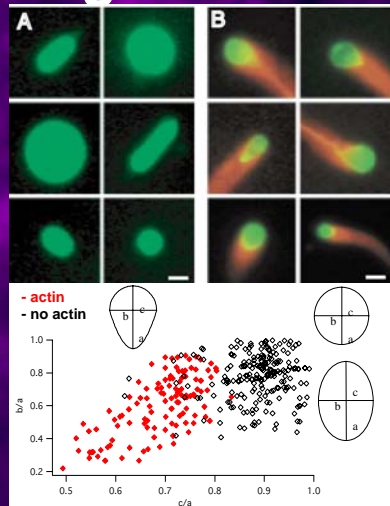


# Movement of ActA-coated beads in cytoplasmic extract

Lisa Cameron



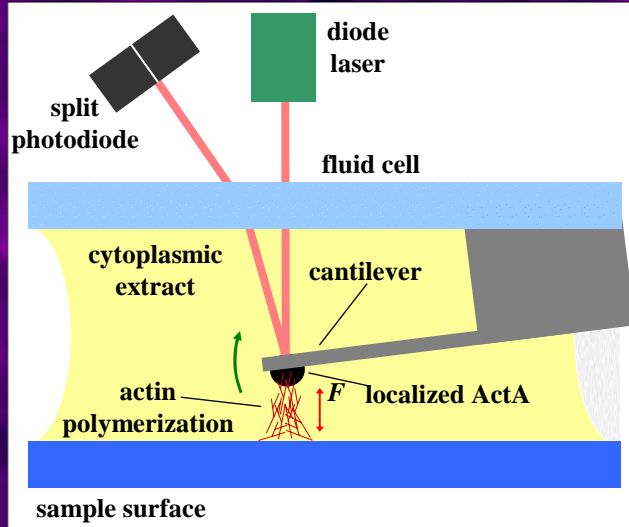
# Force magnitude: 1 comet tail generates ~1 nanoNewton



Vesicle deformation shows that tails squeeze their cargo

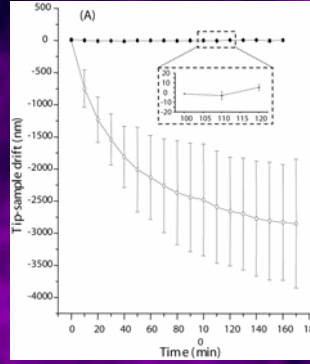
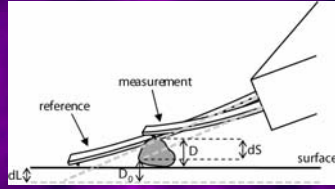
Paula Giardini

### Experimental design for measurement of comet tail force using microfabricated cantilevers



Dan Fletcher

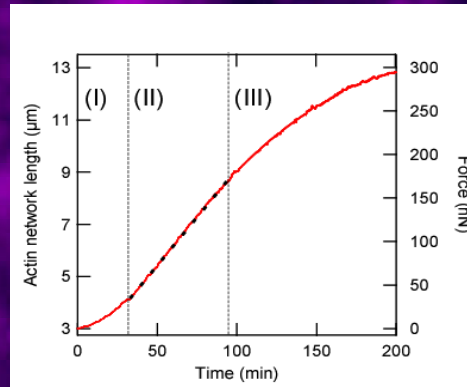
## Dual-beam cantilever AFM design for measuring slow, strong network growth



Drift over 3 hours reduced from 3  $\mu\text{m}$  to a few nm

Dan Fletcher, Jason Choy

## Cantilever deflection driven by actin comet tail growth



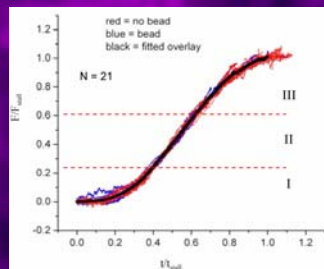
- I. Acceleration
- II. Constant growth
- III. Slowing to stall

NOTE: scale is in NANONEWTONS

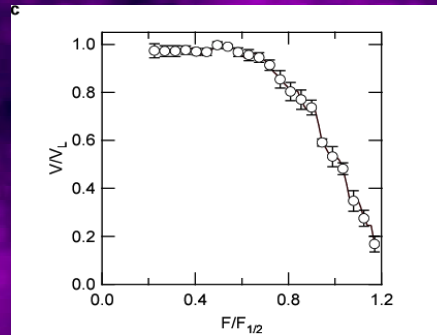
Dan Fletcher, Sapun Parekh

## Force-velocity curve shows distinct phases

Stall force =  
 $2.9 \pm 1.3 \text{ nN}/\mu\text{m}^2$   
 (~7 pN/filament)

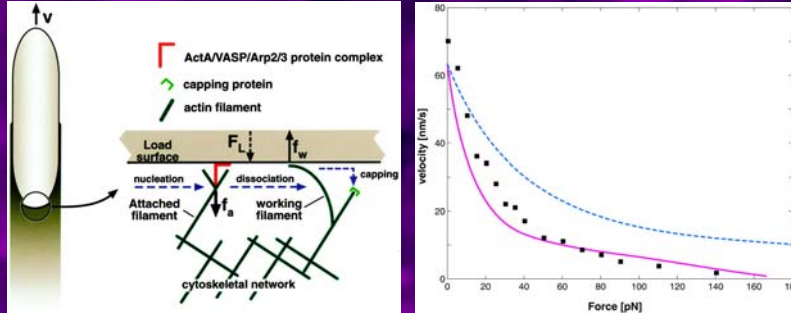


Curve shape is similar  
 regardless of surface geometry

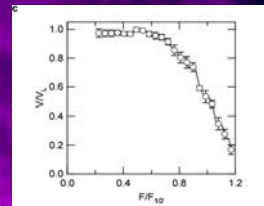


Dan Fletcher, Sapun Parekh

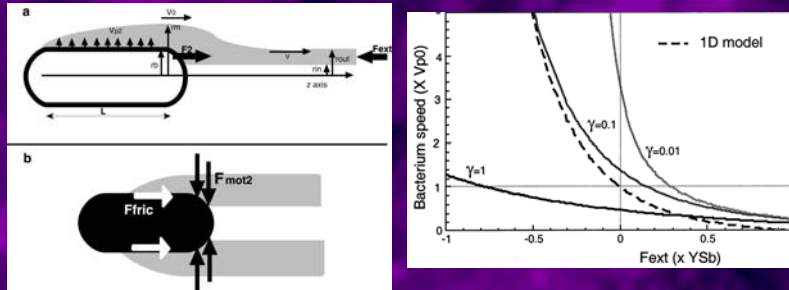
# What did the models predict?



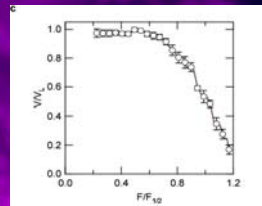
Mogilner and Oster, 2003  
 "Tethered Ratchet"  
 NO flat phase  
 Force decrease is concave up



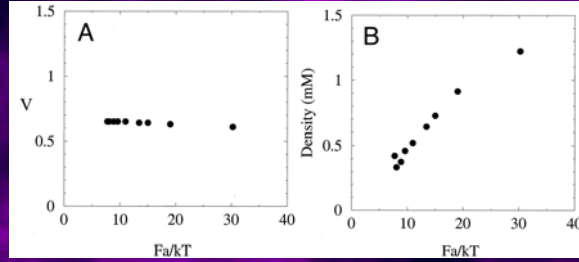
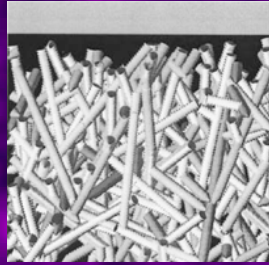
# What did the models predict?



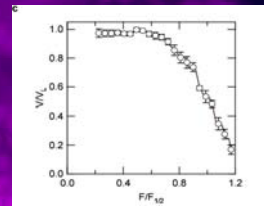
Gerbal, Chaikin, Rabin & Prost 2000  
 "Gel Elastic Model"  
 NO flat phase  
 Force decrease is concave up



## What did the models predict?



Carlsson, 2001  
 "Autocatalytic Nucleation"  
 Flat phase lasts because gel  
 density increases!!  
 Not carried out to fall-off



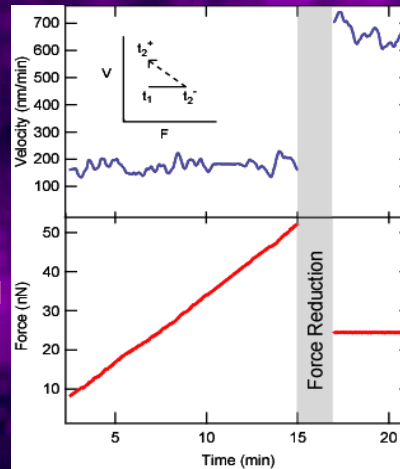


## Hysteresis in the force-velocity curve

Force-loading to pre-stress gel may increase filament density

Restoration to lower load causes FASTER gel growth: up to 3X faster than same load without prestressing

Conclusion: Working out makes you stronger!

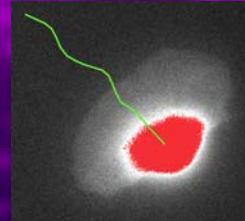


Dan Fletcher, Sapun Parekh

# History-dependent effects in whole-cell movement?

QuickTime™ and a Cinepak decompressor are needed to see this picture.

QuickTime™ and a Cinepak decompressor are needed to see this picture.



**Collisions**

**Oscillations**

**Fish keratocytes:  
Normal persistent motion**

QuickTime™ and a Cinepak decompressor are needed to see this picture.

QuickTime™ and a Animation decompressor are needed to see this picture.

**Symmetry-breaking**

## Summary

**Biological toolkits for mechanical problems  
in dynamic self-organization**

**Actin polymerization-based motility:**

Bacterial movement is stereotyped, geometrically  
simple, molecularly well-defined

Whole-cell movement is the next frontier

**Force generating elements act in groups:**

Spatial arrangement matters (networks > bundles)

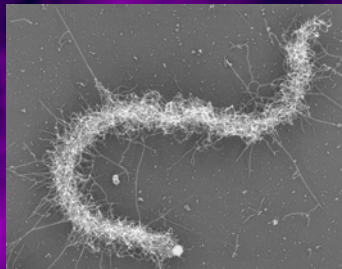
History matters

How are they coordinated in time and space for  
whole cell movement? What about movements  
of cells in complex tissues?

## Acknowledgements

### Current:

Greg Allen, Erin Barnhart, Annie Brotcke, Natalie Dye, Aretha Fiebig, Matthew Footer, Karine Gibbs, Kinneret Keren, Catherine Lacayo, Peter Lee, Zach Pincus, Cyrus Wilson, Patricia Yam



### Recent:

David Baldwin, Lisa Cameron, Martijn van Duijn, Dan Fletcher, Paula Giardini, Susanne Rafelski, Rachael Ream, Jennifer Robbins, Fred Soo, Alexander van Oudenaarden

### Collaborators:

Marileen Dogterom, Jacob Kerssemakers (AMOLF)  
Dan Fletcher, Jason Choy, Allen Liu, Sapun Parekh (UC Berkeley)  
Manuel Amieva, Glen Otto, Mickey Pentecost (Stanford)  
Tom Silhavy, Dan Isaac (Princeton)  
Roger Hendrix, Jun Xu (U. Pittsburgh)  
Alex Mogilner Boris Rubenstein (UC Davis)  
Paul Wiseman, Ben Hebert (McGill)  
Lin Ji, Gaudenz Danuser (Scripps)  
Theresa Harper (Quantum Dots, Inc.)  
Dan Portnoy, Anna Bakardjiev, Pete Lauer, Vicki Auerbuch (UC Berkeley)